

## Tracking Nanocars Using Single Molecule Spectroscopic Methods

#### Introduction

- Nanocars are molecular machines designed to roll over surfaces.
- Rotational motion of the nanocar wheels is thought to result in directional motion.
- The ultimate goal is to use them to transport cargo (which may be as small as a single molecule!) from place to place.



A computer render of the fullerene nanocar, courtesy reference 1.

## Goals

- The goal of this project is to provide a method of optically tracking nanocars.
- Useful to have a quick method of extracting useful information, such as diffusion coefficients and trajectories.
- Also useful to be able to do this in-situ with a non-invasive technique.



A photograph of a section of th laser setup used to excite the fluorescent dyes.



An image from the microscope.

• Compare consecutive frames for movement.

#### Challenges

Frame 1 - Image With Points Found

• Take images (via a scanning confocal microscope), and fit a point spread function to each molecule (allows us to get position beyond the diffraction limit).



- The nanocars are a few nanometers large, however we can only resolve ~260 nm.
- To pinpoint the location of the nanocar more accurately, we use a centroid fit.
- Parameters from this fit are used to eliminate aggregates of single molecules.
- Photoblinking is a reversible phenomenon where the dye molecule will enter into a triplet state, temporarily not emitting any light.
- Photobleaching is a process where the dye undergoes irreversible photochemistry and stops emitting light.



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### Algorithm

In order to detect movement we have to identify molecules in consecutive frames as the same.

# A cartoon showing association.

In the first two instances we can sort out which molecule went where, while in the third we cannot and must ignore both.

- By looking for molecules a certain distance from the ones we see in the first frame we make potential matches.
- If there is more than one match per molecule we can use a pruning technique to remove known matches to eliminate possible matches, allowing us to associate more molecules between frames.
- If we still have more than one match we must remove both, as we don't want wrongly associated molecules.

### Verification

- To make sure the algorithm works we can intentionally shift frames.
- The data below show no shift compared with *every other* frame shifted by 8.486 pixels (6x6). So we expect to see half the molecules showing a shift and half showing no shift.



#### Program

• A MATLAB program was written to perform all of the tests (search radius, histogram, scatterplot, trajectory, and diffusion constant calculation).















#### **Data – Search Radius**

• By varying the size of the radius of the circle that we draw to look for molecules we can vary the number of particles associated.

> Too Small: The second molecule might not be found.

> Optimum: All, or most of the particles are associated.

Too Large: Molecules fall into overlap, and are discarded.

100 frames

The peak association efficency for the nanocars occurs farther out indicating that the molecules are moving a greater amount than the Tritc sample.

## **Data – X-Y Scatter Plots**

Scatterplots can be used to detect drift in the system, as



Search Radius

#### **Data – Radius Histograms**

greater amount for a given association radius size.



Qu, X., Wu, D., Mets, L., Scherer, N. F. Nanometer-localized multiple single-molecule fluorescence microscopy. PNAS. 2004, vol. 101, no. 31. Cheezum, M. K., Walker, W. F., Guilford, W. H., Quantitative Comparison of Algorithms for Tracking Single Fluorescent Particles. Biophys. J. 2001, 81, 2378.

Thompson, R. E., Larson, D. R., Webb, W. W. Precise Nanometer Localization Analysis for Individual Fluorescent Probes. Biophys. J. 2002, 82, 2775. Gimzewski, J. K., Joachim, C. Nanoscale Science of Single Molecules Using Local Probes. Science. 1999, 283, 1683.



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